

A New Cembranolide from the Soft Coral *Sinularia capillosa*

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A new cembranolide, capillolide (**1**), and three known cembranolides were isolated from the soft coral *Sinularia capillosa* collected from the South China Sea. Their structures and the relative stereochemistry of **1** were deduced on the basis of spectroscopic methods.

Soft corals of the genus *Sinularia* yield a wide variety of sesquiterpenes, diterpenes and steroids.^{1–4} Bowden has reported the isolation of 15 furanosesquiterpenes from *Sinularia capillosa* collected at Magnetic Island (Townsville), Australia.⁵ *Sinularia* spp. are very abundant in the South China Sea. In our search for bioactive substances from soft corals, a new cembranolide, capillolide (**1**), and three known compounds (**2–4**) were obtained from *Sinularia capillosa* Tix.-Dur. (Alcyoniidae) collected from the Bay of Sanya, Hainan Island, China.

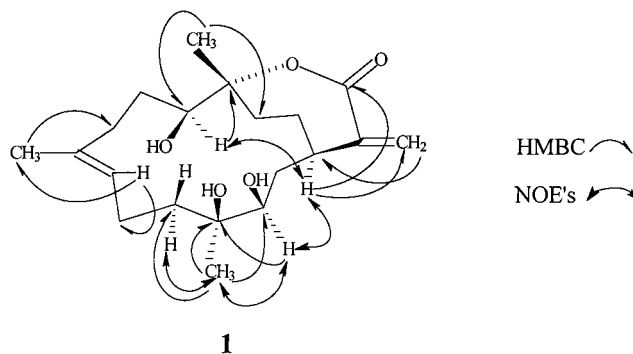
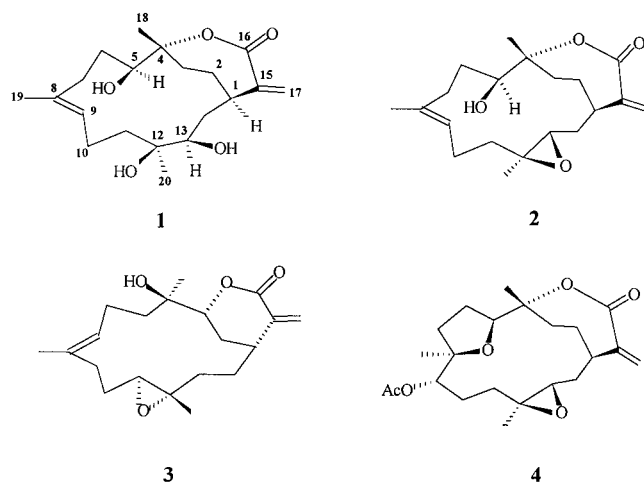


Figure 1. Important ^1H – ^{13}C long-range correlations and NOE interactions of **1**.

secondary and one tertiary hydroxyl groups. Two olefinic carbons were further identified. A signal at δ 5.03 attributed to an olefinic proton, together with a methyl carbon signal at δ 15.9, indicated the *E* configuration for this double bond. Signals for a vinyl methyl at δ 1.48 (3H, s) and two tertiary oxygenated methyl groups at δ 1.31 (3H, s) and 1.22 (3H, s) were observed in the ^1H NMR spectrum. The ^1H NMR and ^{13}C NMR spectra of **1** clearly revealed its cembranolide skeleton. All the ^1H and ^{13}C NMR signals of **1** were unambiguously assigned based on ^1H – ^1H COSY, HMQC, and HMBC NMR experiments. The ^1H – ^{13}C HMBC NMR technique was employed to position the hydroxyl groups about the cembrane ring as illustrated in Figure 1.

The relative stereochemistry of **1** was deduced mainly by 2D NOESY and NOE difference data. In these experiments, both H-13 and H-5 showed NOEs with H-1 but not with H₃-18. Moreover, H₃-20 shared mutual NOE enhancement with H-13 and H-11 α at δ 1.41. These observations indicated that all three hydroxyl groups in **1** were β -oriented and H-1, H-5, H-13, and CH₃-20 were α -oriented with respect to this ring (Figure 1). An additional argument for the stereochemistry of **1** is that **1** and sinulariolide (**2**) were obtained from the same animal, and their NMR spectral data at H₃-18 and C-4 are almost identical (H₃-18 δ 1.31, C-4 δ 87.2 for **1** and H₃-18 δ 1.34, C-4 δ 87.0 for **2**). Furthermore, since H₃-18 did not exhibit a NOE response with H-5, it is reasonable to conclude that CH₃-18 of **1** has the same β -configuration as **2**.

The identities of compounds **2–4** were established by comparison of their spectral data with those of the known compounds reported. They are sinulariolide (**2**),⁶ flexibilide (**3**),⁷ and (1*R*,13*S*,12*S*,9*S*,8*R*,5*S*,4*R*)-9-acetoxy-5,8:12,13-diepoxycebr-15(17)-en-16,4-olide (**4**).⁸

Capillolide (**1**) showed moderate cytotoxic activity. ED₅₀ values in a test against P-388 and L1210 cell lines are 15.0 and 18.5 $\mu\text{g}/\text{mL}$, respectively. ED₅₀ values for cytotoxicity toward the P388 and L1210 cell lines are 8.5 and 10.0 $\mu\text{g}/\text{mL}$.

Capillolide (**1**) was isolated as colorless crystals. Its molecular formula, C₂₀H₃₂O₅, was established by FABMS m/z 375 [M + Na]⁺ and ^{13}C NMR data implying five degrees of unsaturation. Its IR spectrum showed a broad absorption between 3000 and 3400 cm⁻¹ (OH stretching) and a strong absorption at 1700 cm⁻¹, consistent with the presence of an α,β -unsaturated ester function. The ^{13}C NMR and DEPT experiments revealed the presence of one carbonyl carbon, two double bonds, and four oxygenated carbons. ^1H and ^{13}C NMR spectra, combined with 2D NMR experiments, allowed the chemical structure of **1** to be established.

The presence in the ^1H NMR spectrum of two broad singlets at δ 6.37 (1H, br s) and 5.75 (1H, br s) suggested the presence of an exocyclic α -methylene lactone ring. This was supported by the ^{13}C NMR (δ 169.7, 142.2, 125.0) and IR spectra (1700 cm⁻¹). A tertiary oxygenated carbon signal at δ 87.2 suggested the presence of an ϵ -lactone moiety,¹ which was confirmed by the ^1H – ^1H COSY and HMBC experiment (Table 1 and Figure 1). Carbon signals were observed at δ 68.4 (d), 73.3 (d), and 74.9 (s), and two proton signals at δ 4.16 and 3.38 indicated the presence of two

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Table 1. ^1H and ^{13}C NMR Signals of Capillolide (1)

position	^{13}C ^a	^1H (J) ^b	^1H - ^1H COSY	HMBC
1	36.3 d	2.35, m	2.12, 1.62, 2.07, 1.21	6.37, 5.75, 2.07, 1.21
2	36.2 t	2.07, m 1.21, m	2.35, 1.21, 1.78, 2.06 2.35, 2.07, 1.78, 2.06	1.78, 2.06
3	31.6 t	1.78, m 2.06, m	2.07, 2.06, 1.21 2.07, 1.78, 1.21	2.07, 1.21
4	87.2 s			1.78, 2.06, 4.16, 1.31
5	68.4 d	4.16, dd(7.3, 1.1 Hz)	1.39, 2.18	1.78, 2.06, 1.31
6	30.2 t	1.39, m 2.18, m	4.16, 2.18, 1.65, 2.10 4.16, 1.39, 2.10	1.65, 2.10
7	36.6 t	1.65, m 2.10, m	1.39, 2.10 1.39, 1.65, 2.18	1.48
8	135.6 s			1.48, 5.03
9	127.2 d	5.03, dd(10.9, 1.2 Hz)	1.68, 2.09	1.48, 1.68, 2.09
10	22.1 t	1.68, m 2.09, 1.90	5.03, 2.09, 1.90 1.68, 1.90, 1.41	5.03
11	37.9 t	1.90, m 1.41, m	1.68, 2.09, 1.41 2.09, 1.90, 1.68	1.22
12	74.9 s			1.90, 1.41, 1.22, 3.38
13	73.3 d	3.38, dd(7.3, 1.1 Hz)	2.12, 1.62	1.22, 2.12, 1.62
14	28.6 t	2.12, m 1.62, m	1.62, 2.35 2.12, 2.35	2.35
15	142.2 s			2.35
16	169.2 s			2.35, 6.37, 5.75
17	125.0 t	6.37, br s 5.75, br s		2.35
18	22.7 q	1.31, s		4.16
19	15.9 q	1.48, s		1.65, 2.10
20	24.1 q	1.22, s		

^a ^{13}C NMR determined at 125 MHz in CDCl_3 . Multiplicities determined by DEPT and HMQC experiments. ^b ^1H NMR determined at 500 MHz in CDCl_3 .

mL for sinulariolide (2), 1.5 and 3.0 $\mu\text{g}/\text{mL}$ for flexibililide (3), and 2.5 and 5.0 $\mu\text{g}/\text{mL}$ for (1*R*,13*S*,12*S*,9*S*,8*R*,5*S*,4*R*)-9-acetoxy-5,8:12,13-diepoxycebr-15(17)-en-16,4-olide (4).

Experimental Section

General Experimental Procedures. The melting points were determined using a X₄ micro-melting point apparatus and are uncorrected. IR spectra were recorded with a EQUINOX55 (Bruker) spectrophotometer. ^1H and ^{13}C NMR spectra were recorded with a Varian Unity INOVA spectrophotometer at 500 and 125 MHz, respectively, in CDCl_3 using TMS as internal standard. MS spectra were obtained with a VG ZAB-MS mass spectrometer. Optical rotations were measured on a Perkin-Elmer 341 polarimeter. Si gel (Qingdao, 200–300 μ) was used for column chromatography.

Animal Material. The soft coral *Sinularia capillosa* (Alcyoniidae) was collected off the Bay of Sanya, Hainan Island, China. A voucher specimen (No. 98-sy-21) is preserved in the Research Centre of Organic Natural Products, Zhongshan University.

Extraction. The soft coral *S. capillosa* (1.2 kg, dried wt) was extracted with EtOH. The EtOH extract was concentrated in vacuo, and the residue was extracted successively with petroleum ether, CH_2Cl_2 , and n-BuOH. The CH_2Cl_2 extract was subjected to Si gel column chromatography eluting with petroleum ether containing increasing concentrations of EtOAc. Four fractions were obtained: Fraction A (fr. 26–28) gave sinulariolide (2) (15 mg), fraction B (fr. 29,30) gave compound (4) (20 mg), fraction C (fr. 50–53) yielded flexibililide (3) (100 mg), and fraction D (fr. 79,80) afforded capillolide (1) (500 mg).

Capillolide (1): colorless crystals; mp 158–160 °C; $[\alpha]_D^{25} +42.8^\circ$ (c 0.05, EtOH); IR (KBr) ν_{max} 3417, 2930, 1700, 1618, 1049 cm^{-1} ; ^1H NMR and ^{13}C NMR, see Table 1; FABMS m/z 375 $[\text{M} + \text{Na}]^+$; EIMS m/z 335 $[\text{M} + 1 - \text{H}_2\text{O}]^+$ (37), 317 (23), 299 (16), 279 (10), 154 (64), 140 (62), 121 (35), 55(100).

Sinulariolide (2): colorless crystals, mp 171–173 °C; $[\alpha]_D^{25} +134.4^\circ$ (c 0.03, EtOH); IR (KBr) ν_{max} 3456, 2937, 1712, 1462, 1382, 1238, 1081 cm^{-1} ; ^1H NMR (CDCl_3) δ 6.29 (1H, s; C₁₇-H), 5.42 (1H, s; C₁₇-H), 5.13 (1H, br d, $J = 9.0$ Hz; H-9), 4.08 (1H, dd, $J = 10.0, 7.0$ Hz; H-5), 2.93 (1H, dd, $J = 11.0, 3.0$ Hz; H-13), 1.61 (3H, s; H-19), 1.34 (3H, s), 1.20 (3H, s); ^{13}C NMR

(CDCl_3) δ 168.9 (C-16), 144.4 (C-15), 134.8 (C-8), 126.6 (C-9), 124.1 (C-17), 87.0 (C-4), 68.7 (C-5), 63.9 (C-13), 60.4 (C-12), 38.1 (CH₂), 35.9 (CH₂), 35.1 (C-1), 33.1 (CH₂), 31.7 (CH₂), 31.4 (CH₂), 26.9 (CH₂), 25.0 (CH₂), 22.8 (CH₃), 16.0 (CH₃), 15.8 (CH₃).

Flexibililide (3): colorless crystals, mp 153–155 °C; $[\alpha]_D^{25} -90.4^\circ$ (c 0.03, EtOH); IR (KBr) ν_{max} 3395, 2940, 1731, 1286, 1180, 1030 cm^{-1} ; ^1H NMR (CDCl_3) δ 6.48 (1H, d, $J = 2.0$ Hz; H-17), 5.70 (1H, d, $J = 2.0$ Hz; H-17), 5.24 (1H, br t, $J = 8.0$ Hz; H-7), 3.98 (1H, d, $J = 11.0$ Hz; H-3), 2.81 (1H, dd, $J = 9.0, 4.0$ Hz; H-11), 1.67 (3H, s; H-19), 1.45 (3H, s), 1.32 (3H, s); ^{13}C NMR (CDCl_3) δ 167.1 (C-16), 140.1 (C-15), 134.5 (C-7), 127.8 (C-17), 125.6 (C-8), 82.7 (C-3), 74.1 (C-4), 62.9 (C-11), 58.9 (C-12), 38.6 (CH₂), 35.9 (CH₂), 33.8 (C-1), 33.0 (CH₂), 27.8 (CH₂), 25.3 (CH₂), 24.9 (CH₂), 22.6 (CH₂), 15.4 (CH₃), 15.4 (CH₃).

(1*R*,13*S*,12*S*,9*S*,8*R*,5*S*,4*R*)-9-Acetoxy-5,8:12,13-diepoxycebr-15(17)-en-16,4-olide (4): colorless crystals, mp 148–149 °C; $[\alpha]_D^{25} +39.1^\circ$ (c 0.03, EtOH); IR (KBr) ν_{max} 3440, 2979, 1738, 1715, 1628, 1376, 1236, 1059 cm^{-1} ; ^1H NMR(CDCl_3) δ 6.27 (1H, s; H-17), 5.44 (1H, s, H-17), 5.17 (1H, d, $J = 9.0$ Hz), 4.38 (1H, dd, $J = 9.7$ Hz, 3.0 Hz, H-5), 3.39 (1H, dd, $J = 11.0, 4.0$ Hz; H-13), 3.22 (1H, m), 2.57 (1H, dd, $J = 13.0, 6.5$ Hz), 2.06 (3H, s), 1.25 (3H, s), 1.24 (3H, s), 1.16 (3H, s); ^{13}C NMR- (CDCl_3) δ 171.1, 169.5 (C-16), 145.4 (C-15), 123.5 (C-17), 88.1 (C-4), 85.4 (C-8), 83.4 (C-5), 77.9 (C-9), 60.8 (C-13), 59.9 (C-12), 35.7 (C-1), 34.9 (CH₂), 34.3 (CH₂), 33.6 (CH₂), 33.2 (CH₂), 29.9 (CH₂), 29.3 (CH₃), 27.3 (CH₂), 26.8 (CH₂), 21.0 (CH₃), 17.0 (CH₃), 16.4 (CH₃).

Cytotoxicity Testing. P-388 and L1210 tumor cells were obtained from the Cancer Institute, Sun Yat-sen University of Medical Sciences. Cytotoxicity assays of the tested compounds 1–4 were carried out by a modification of the MTT colorimetric method.⁹ The culture cells were treated at eight concentrations of pure test compounds ranging from 0.00064 to 50 $\mu\text{g}/\text{mL}$. All assays were performed in triplicate. The results were expressed as a percentage, relative to control incubations, and the effective dose required to inhibit cell growth by 50% (ED₅₀) was determined.

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